

# Antimicrobial resistance, adhesion and biofilm formation ability of Salmonella strains isolated from the French pig and pork industry

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# Introduction

Salmonella remains the most frequently detected causative agent in the foodborne outbreaks reported in 2013 in European Union (22.5 % of total outbreaks) (EFSA, 2015). Pork and products thereof are commonly implicated in Salmonella outbreaks. Salmonella Derby and Salmonella Typhimurium are the two main prevalent serovars in the French pig and pork industry.

This study aims at characterizing a collection of S. Typhimurium and S. Derby isolated along the pork meat production chain in France for various phenotypic traits i.e. antimicrobial resistance patterns, motility, adhesion and biofilm formation on abiotic surfaces.

## **Material & Methods**

#### > Bacterial strains

■ 16 S. Typhimurium (including the ATCC® 14028 strain) and 15 S. Derby strains, isolated along the pork meat production chain in France

#### > Motility tests

 Twitching, swarming and swimming motilities were evaluated using nutritive media with 1%, 0.5% (plus 0.5% glucose) and 0.25% agar concentration, respectively.

#### Adhesion and biofilm formation

- Adhesion ability to two abiotic surfaces, i.e. and high-density stainless steel (SS) polyethylene (HDPE), was evaluated after 3hincubation in physiological salt solution at 37°C (ACTIA, 2015).
- Biofilm formation was determined by growing the bacterial cultures for 48 h in 96-well microtiter plates followed by confocal laser scanning microscopy observations.

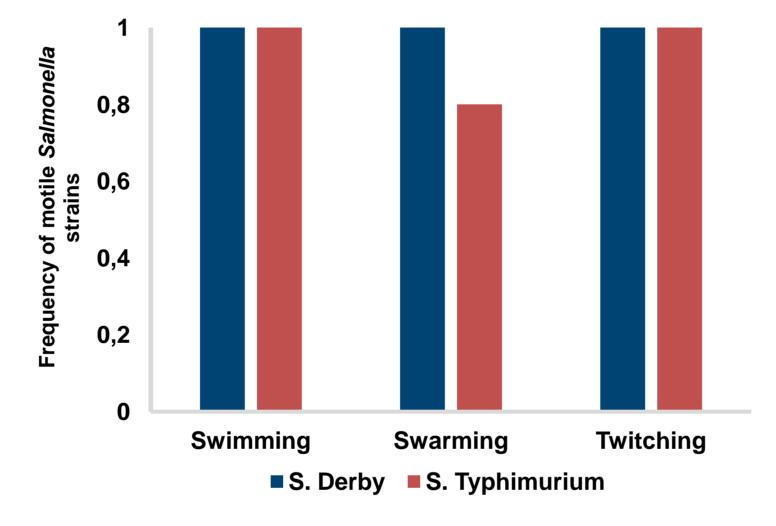
#### > Antimicrobial resistance patterns

- Antibiotic susceptibility tests were performed using a standard microdilution method with the Sensititre® system on EUMVS-2 microtiter plates (Trek Diagnostic Systems, England).
- Susceptibility to biocides commonly found in disinfectant formulations in the French pork industry, was evaluated using a standard microdilution method (Soumet et al., 2005).

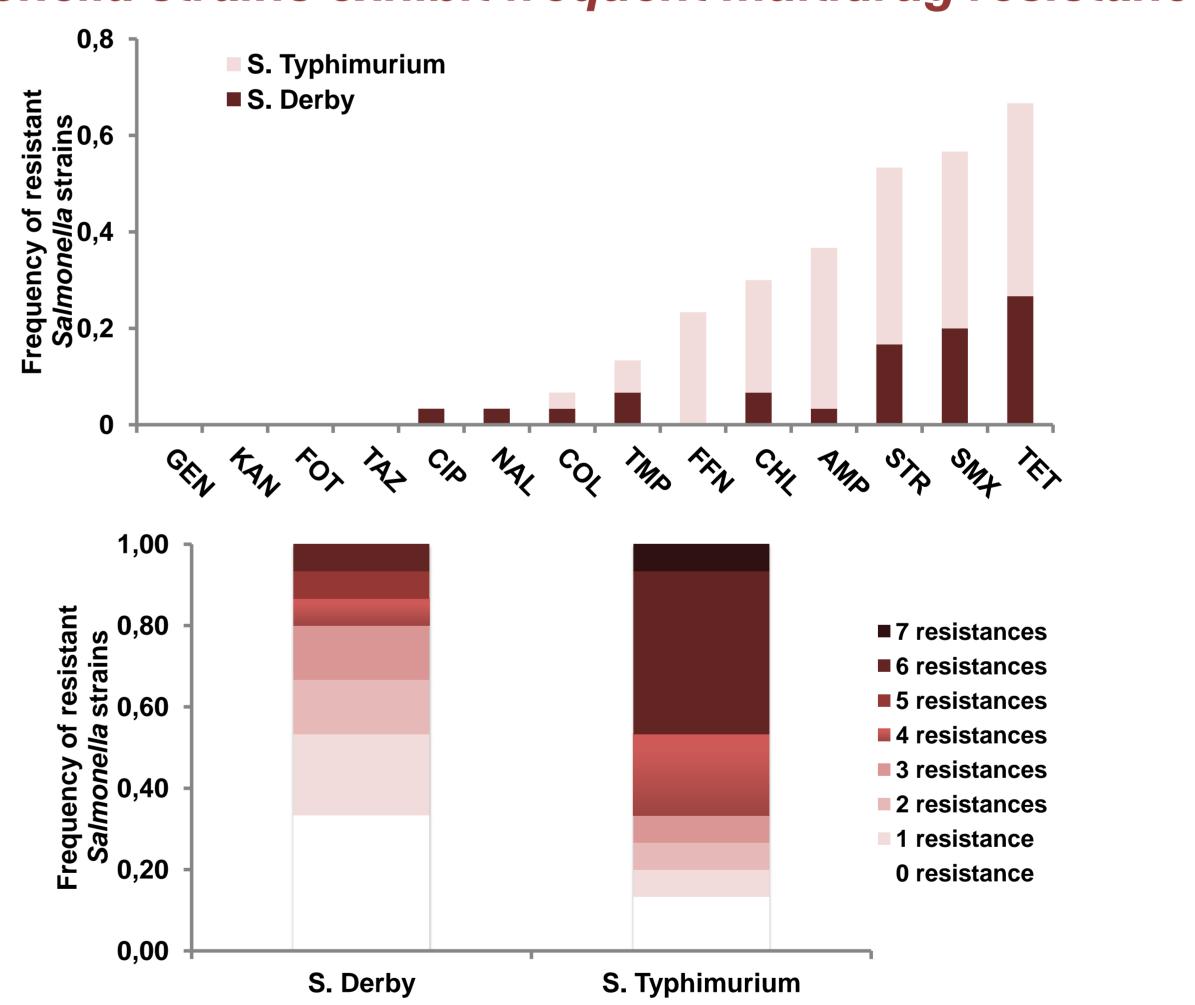
# Results

The majority of Salmonella officient strains displayed efficient motility

Figure 1: Frequency of S. Typhimurium Derby strains displaying active swimming, swarming and twitching motility



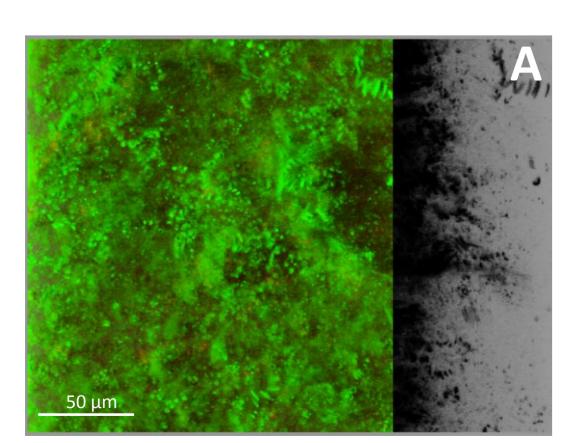
### > Salmonella strains exhibit frequent multidrug resistances



**Figure 4:** Frequency of antibiotic resistances in *S.* Typhimurium and *S.* Derby strains GEN: gentamycin, KAN: kanamycin, FOT: cefotaxime, TAZ: ceftazidine, CIP: ciprofloxacine, NAL: nalidixic acid, COL: colistin, TMP: triméthoprime, FFN: florfenicol, CHL: chloramphenicol, AMP: ampicilline, STR: streptomycin, SMX: sulfamethoxazole, TET: tetracyclin

#### Figure 2: Levels of adherent Salmonella cells according to (A) the serovar Derby and Typhimurium; (B) the type of surfaces SS and PEHD (non-polar surfaces with $\theta_{water} = 95$ ). Levels obtained for the ATCC® 14028 are displayed in red and blue for SS and PEHD, respectively.

> High ability to adhere and form biofilm on abiotic surfaces



S. Derby - S. Typhimurium

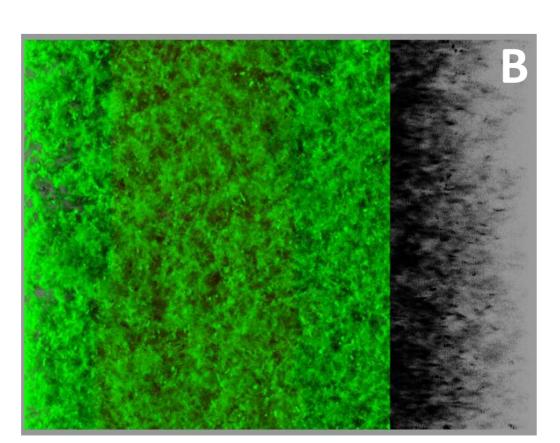


Figure 3: Confocal laser scanning microscopy observations of Salmonella biofilms on 96-well microplate (A: S. Typhimurium strain; B: S. Derby strain)

**Table 1 :** Biocide susceptibility of *S.* Typhimurium and *S.* Derby strains Didecyl Dimethyl Ammonium Chloride (DDAC), benzalkonium chloride (BC), hydrogen peroxide (H2O2), sodium hypochlorite (NaClO), peracetic acid (C<sub>2</sub>H<sub>4</sub>O<sub>3</sub>), Galox Horizon (end-use formulation with DDAC and glutaraldehyde)

Tolerance thresholds ( $\mu$ g/ml): DDAC > 6, Galox Horizon > 4, BC > 32, H<sub>2</sub>O<sub>2</sub> > 40, NaClO > 600, C<sub>2</sub>H<sub>4</sub>O<sub>3</sub> > 0.015

Salmonella strains (n = 30)	DDAC	Galox Horizon	ВС	H <sub>2</sub> O <sub>2</sub>	NaCIO	Peracetic acid
CMI (µg/ml)	4-8	3-6	24-64	30-40	300-600	0.015
<i>n</i> Tolerant/ <i>n</i> Total	6/30	11/30	1/30	0/30	0/30	0/30
ATCC 14028 CMI (µg/mI)	1.5	1.5	12	30	400	0.015

# Conclusion



Other studies are necessary to increase our understanding of the determinants and mechanisms involved in the predominance and persistence of these two specific serovars in pigs



Références ACTIA, RMT CHLEAN (2015) Guide pratique pour la réalisation d'essais d'adhésion microbienne er laboratoire, une étape préliminaire à la formation de biofilms EFSA and ECDC (European Food Safety Authority and European Centre for Disease Prevention and Control) (2015). The European Union Summary Report on Trends and Sources of Zoonoses, Zoonotic Agents and Food-borne Outbreaks in 2013. EFSA Journal 2015; 13(1):3991, 162 pp. doi:10.2903/j.efsa.2015.3991 Soumet, C., Ragimbeau, C. & Maris, P. 2005. Screening of benzalkonium chloride resistance in *Listeria* monocytogenes strains isolated during cold smoked fish production. Lett Appl Microbiol, 41, 291-6